

## A Brief Overview on Early Events of Xa21 Mediated Pattern Triggered Immunity

M H M Mubassir\*<sup>1</sup>, M Abu Naser<sup>2</sup>, Mohd Firdaus Abdul-Wahab<sup>3</sup>, Salehuddin Hamdan<sup>3</sup>

<sup>1</sup>Department of Mathematics and Natural Sciences, BRAC University, Bangladesh

<sup>2</sup>Scientific Computing Solutions, Bangladesh

<sup>3</sup>Faculty Bioscience and Medical Engineering, Universiti Teknologi Malaysia, Malaysia

\*Corresponding author: E-Mail: [mhmmubassir@bracu.ac.bd](mailto:mhmmubassir@bracu.ac.bd), Cell: +8801711130886

### ABSTRACT

In plant associated pattern-triggered immunity (PTI), pattern-recognition receptors (PRRs) play crucial role in the mechanism of first line of defense against pathogenic invasion. Throughout the event, PRR binds with microorganism's pathogen-associated molecular pattern (PAMP) and recruit co-receptor protein to initiate the primary defense signal. Although several plant PRRs have been identified, very few of them have been fully characterized and studied so far. PRR Xa21 perceives PAMP RaxX21 and activates pattern triggered immunity in *Oryzae* spp (rice). The early events regulated by Xa21 is mainly binding of it with PAMP RaxX21 as well as recruit a co-receptor protein OsSerk2. This review article elucidates the steps of activating first line of defense mechanism of rice plant arbitrated by PRR Xa21.

**KEY WORDS:** PRR, PAMP, PTI, Xa21, Defense

### 1. INTRODUCTION

The first layer of defense mechanism in plant is referred as pattern triggered immunity (PTI) (Jones, 2006; Ronald, 2010; Tsuda, 2009). Plant has many immune receptors which are localized at cell surface. These receptors play a vital role in PTI. These receptors perceive pathogen associated molecular pattern (PAMPs) or microbial associated molecular pattern (MAMPs) secreted by microorganisms like bacteria, fungi etc. These PRRs are mainly two types. One is receptor like kinase (RLK) which has kinase domain at the end and another one is receptor like protein (RLP) which does not have any kinase domain (Zipfel, 2014). Receptor like kinase has four main region which are leucine rich repeat (LRR), a single pass transmembrane domain (TM), one juxtamembrane domain (JM) and an intracellular kinase domain (Song, 1995). The LRR binds with PAMPs/MAMPs and through the TM and JM the signal is transferred to the inner side of the cell by kinase domain and PTI is activated. During this event a co-receptor protein is recruited which is required for the full activation of PTI (Chen, 2014).

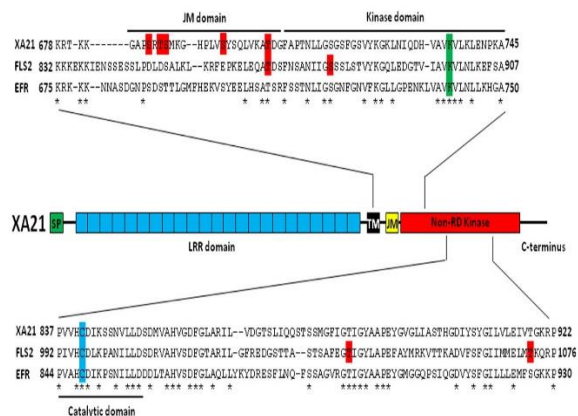
Successful pathogens can avoid the pattern triggered immunity and secrets effectors by its type 3 secretion system (Thomma, 2011). Plant can also avoid harmful activities caused by effectors by its resistant proteins R. Most of these proteins are intracellular receptor proteins of the nucleotide binding leucine rich repeat (NB-LRR). Effector triggered immunity occur more rapidly than pattern triggered immunity (Tao, 2003; Tsuda, 2010). These effectors previously known as avirulence factors (Chisholm, 2006; Bent, 2007). This ETI is also known as gene-for-gene hypothesis where both gene product from plant and pathogen interacts with each other in a receptor-ligand manner (Schurch, 2004).

Rice Xa21 is a of PRR-RK falls under the XII sub-family (LRR-RK XII) with the other two best characterized PRR-RKs *Arabidopsis* FLS2 and EFR (Holton, 2015). Sub-family XII is one of the most expanded subfamily of LRR-RK in rice which encodes mostly for PRRs or PRR-associated RKs due to presence of non-RD kinase domains (Dardick, 2006).

Due to the presence of Xa21, rice plant shows robust resistance to bacterial leaf blight (BLB) causing *Xanthomonas oryzae* pv. *Oryzae* (*Xoo*) (Wang, 1996; Ishiyama, 1922). BLB is regarded as one of the major diseases of rice (Swings, 1990), which can cause yield loss up to 70% (Mew, 2001). The best cost effective control of this disease is achieved by using of resistant cultivars (Nino-Liu, 2006; Pinta, 2013). This mini overview elucidates the initial steps of activating front line of defense mechanism arbitrated by PRR Xa21.

#### **Brief overview of Pti Arbitrated by XA21:**

**Domains of Xa21:** Xa21 has 23 leucine rich repeat (LRR) domains (Figure.1), one transmembrane (TM) domain, one juxtamembrane domain (JM) and one kinase domain.

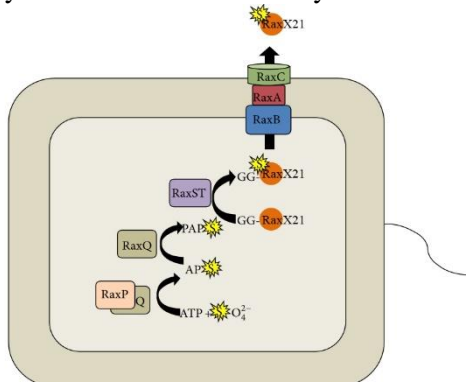


**Figure.1. Detailed domain structures of Xa21 (Park, 2012)**

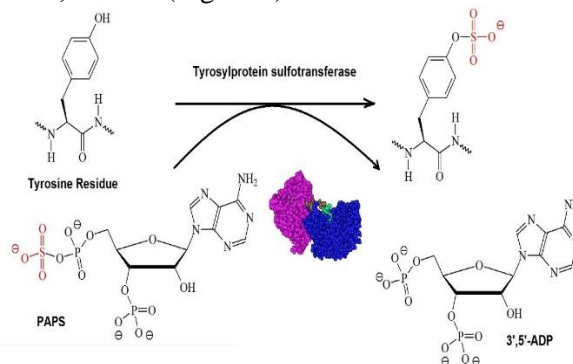
LRR domain which is localized outside of the plant membrane, binds to bacterial peptide RaxX21 (Pruitt, 2015) where JM domain transfers the signal to kinase domain. This activates the PTI where co-receptor protein OsSerk2 associates with Xa21 helping to trigger the plant defense mechanism. Perhaps due to this, the in situ structural complexity of the whole protein, Xa21 structure is yet to be determined experimentally.

**RaxX21-sY as activator of Xa21 mediated immunity:** For the activation of Xa21 mediated immunity, six *Xoo* genes of two functional classes are required. RaxA, raxB and raxC which belongs to the first class, encode a membrane fusion protein (MFP), an adenosine triphosphate (ATP) binding cassette (ABC) transporter protein, and an outer membrane protein (OMP), respectively. These all together are predicted to comprise a type 1 secretion system. *Xoo* Bacteria secretes RaxX21 through this type 1 secretion system (Figure.2) (Ronald, 2014). But before the secretion, this RaxX21 become sulphated with the help of another second class of rax genes raxP and raxQ (Shen, 2002). These encode an ATP sulfurylase and an adenosine 5'-phosphosulphate (APS) kinase and function in concert to produce 3'-phosphoadenosine 5'-phosphosulphate (PAPS), the universal sulfuryl group donor (Y shen 2002) (Figure.3).

This sulphation takes place at the 7<sup>th</sup> amino acid tyrosine of RaxX21. It is predicted that about 1% of all tyrosines are sulphated in eukaryotic cells where this sulphation plays a vital role as mediator in protein-protein interaction. The enzymes tyrosylprotein sulfotransferases or TPSTs catalyze the transfer of a sulfonate group from the donor compound 3'-phosphoadenosine 5'-phosphosulfate or PAPS to the hydroxyl group of a lumenally oriented peptidyltyrosine residue to form a tyrosine O4-sulfate ester and 3',5'-ADP (Figure.3).



**Figure.2. Synthesis and secretion of RaxX21 protein (Ronald, 2014)**



**Figure.3. The tyrosylprotein sulfotransferase mechanism in RaxX21 7<sup>th</sup> AA (Biosynthesis, 2015)**

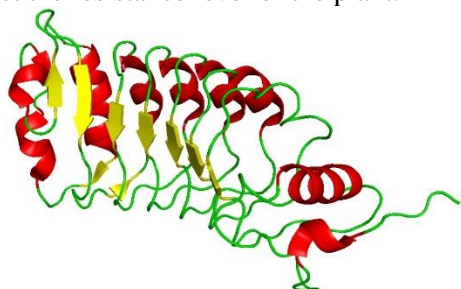
**Xa21 Leucine rich repeat domain recognize RaxX21-sY:** Immediately after secretion of RaxX21-sY (sulphated RaxX21) by *Xoo* through type 1 secretion system, it is recognized by rice PRR Xa21 leucine rich repeat domain. Xa21, not having the transmembrane and kinase domains, shows partial resistance to bacterial blight. This proves that when RaxX21 binds with LRR of Xa21, in absence of transmembrane and kinase domain, an unidentified receptor kinase is activated which transduce the signal of defense mechanism which leads the plant to show partial resistance.

**OsSerk2 Co-receptor regulates the Xa21 mediated immunity:** In case of FLS2 and EFR mediated immunity BAK1 co-receptor protein positively regulates the innate immunity of the plant. When PAMP binds with FLS2 or EFR, it forms an instantaneous complex with BAK1. Like FLS2 and EFR, Xa21 and OsSerk2, a co-receptor protein (Figure.4) for Xa21 mediated immunity form a heterodimer complex upon recognition of RaxX21-sY.

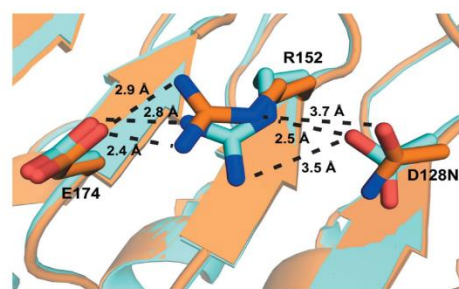
Moreover, in case of FLS2 mediated immunity, the PAMP flg22 acts as a molecular glue by stabilizing the interaction between receptor FLS2 and co-receptor BAK1 (Sun, 2013). But in the case of Xa21 mediated immunity,

the strong association between Xa21 and co-receptor OsSerk2 without ligand treatment in mature rice plant proves that the association between Xa21 and OsSerk2 is PAMP independent.

Again, mutation in BAK1 Asp122 amino acid has altered interaction with the receptor FLS2 suggests that mutation in co-receptor protein OsSerk2 may have significant impact on its binding with Xa21. Later crystallographic structure of the OsSerk2 LRR domain and its mutant version (D128) proved that the mutation has significant impact on its intra-protein binding. In the wild type, the OsSerk2 Asp128 forms a salt bridge to Arg152 (McAndrew, 2014). But in mutant, Arg152 forms a salt bridge with the nearby residue Glu174 (Figure.5). This change in salt bridge formation may have significant impact on binding of the protein with its PRR Xa21 and thus may affect the resistance level of the plant.



**Figure.4. Cartoon representation of OsSerk2 structure (PDB ID: 4Q3G) (McAndrew, 2014)**



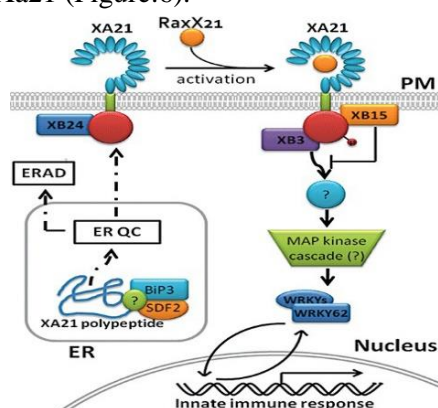
**Figure.5. Salt bridge formation by OsSerk2 Arg152 with other residues (McAndrew, 2014)**

**JM domain and other proteins regulate Xa21 mediated immunity:** JM domain of Xa21 protein plays vital role in regulation of Xa21 mediated immunity. Autophosphorylation of the residue Ser686, Thr688 and Ser689 has vital role in case of stabilizing the Xa21 protein. Mutation in any of this three amino acids show partial resistance to bacterial blight disease compared to wild type Xa21 (Xu, 2006). Also Thr705 in the JM region is required for binding of Xa21 protein with other regulatory protein XB3, XB10, XB15, XB24 (Chen, 2010).

Ring finger ubiquitin ligase XB3, which is trans phosphorylated by kinase domain of Xa21 is required for Xa21 mediated immunity (Wang, 2010). Moreover XB10, a WRKY transcriptional factor, can interact with kinase domain of Xa21 and can negatively regulate the Xa21 mediated immunity (Park, 2010).

Overexpression of XB10, a WRKY transcriptional factor suppress the expression of Xa21 mediated immunity and this negatively regulate the PTI mechanism in rice plant (Peng, 2008). Like XB10, another protein phosphatase XB15 negatively regulates the Xa21 mediated immunity (Park, 2008).

XB24 another ATPase interacts with Xa21 and regulates the immunity mediated by Xa21 (Chen, 2006). XB24 keeps Xa21 in dormant stage by promoting auto phosphorylation of certain Ser/Thr sites while physically associated with it. Upon recognition of RaxX21, XB24 dissociates from Xa21 and PTI mechanism is activated. Thus XB24 regulates the Xa21 before its interaction with RaxX21 while XB3, XB10 and XB15 controls the activity of Xa21 after recognition of RaxX21 by Xa21 (Figure.6).



**Figure.6. An overview of Xa21 mediated immunity (Ronald and Beutler, 2010)**

## 2. CONCLUSION

Much of what has been experimented above describes an overview of PRR Xa21 along with its perception of bacterial PAMP RaxX21 and OsSerk2. Since, Xa21 has been shown to play a key role in pattern triggered immunity, clear understanding of its structural details is necessary before concluding its functional role in the disease defense mechanism. Future research can be carried out to retrieve the crystallographic structure of Xa21 and its interaction with RaxX21-sY as well as co-receptor OsSerk2 to deduce its functional detail on heterodimer PTI complex formation.

## REFERENCES

- Bent A.F and Mackey D, Elicitors, effectors, and R genes, the new paradigm and a lifetime supply of questions, *Annu. Rev. Phytopathol.*, 45, 2007, 399-436.
- Biosynthesis, Sulfated Tyrosine Peptides or Sulfotyrosine Peptides, 2015.
- Chen X, An ATPase promotes autophosphorylation of the pattern recognition receptor XA21 and inhibits XA21-mediated immunity. *Proceedings of the National Academy of Sciences*, 107 (17), 2010, 8029-8034.
- Chen X, An XA21-associated kinase (OsSERK2) regulates immunity mediated by the XA21 and XA3 immune receptors. *Molecular plant*, 7 (5), 2014, 874-892.
- Chisholm S.T, Host-microbe interactions: shaping the evolution of the plant immune response, *Cell*, 124 (4), 2006, 803-814.
- Dangl J.L and Jones J.D, Plant pathogens and integrated defense responses to infection, *nature*, 411 (6839), 2001, 826-833.
- Dardick C and Ronald P, Plant and animal pathogen recognition receptors signal through non-RD kinases. *PLoS pathogens*, 2 (1), 2006, 2.
- Holton N, The phylogenetically-related pattern recognition receptors EFR and XA21 recruit similar immune signaling components in monocots and dicots, *PLoS pathogens*, 11 (1), 2015.
- Ishiyama S, Studies of bacterial leaf blight of rice, Report of the Imperial Agricultural Station, Nishigahara (Konosu), 45, 1922, 233-261.
- Jones J.D and Dangl J.L, The plant immune system, *Nature*, 444 (7117), 2006, 323-329.
- McAndrew R, Structure of the OsSERK2 leucine-rich repeat extracellular domain. *Acta Crystallographica Section D: Biological Crystallography*, 70 (11), 2014, 3080-3086.
- Mew T and Vera Cruz C, Bacterial blight of rice, *Encyclopaedia of plant pathology*, Wiley, New York, 2001, 71-74.
- Nino-Liu D.O, Ronald P.C and Bogdanove A.J, *Xanthomonas oryzae* pathovars, model pathogens of a model crop, *Molecular Plant Pathology*, 7 (5), 2006, 303-324.
- Park C.J, Caddell D.F and Ronald P.C, Protein phosphorylation in plant immunity, insights into the regulation of pattern recognition receptor-mediated signaling, *Frontiers in plant science*, 3, 2012, 177.
- Park C.J, Elucidation of XA21-mediated innate immunity, *Cellular microbiology*, 12 (8), 2010, 1017-1025.
- Park C.J, Rice XB15, a protein phosphatase 2C, negatively regulates cell death and XA21-mediated innate immunity, *PLoS Biol*, 6 (9), 2008, 231.
- Peng Y, OsWRKY62 is a negative regulator of basal and Xa21-mediated defense against *Xanthomonas oryzae* pv. *oryzae* in rice, *Molecular plant*, 1 (3), 2008, 446-458.
- Pinta W, Pyramiding of blast and bacterial leaf blight resistance genes into rice cultivar RD6 using marker assisted selection. *African Journal of Biotechnology*, 12 (28), 2013, 4432-4438.
- Pruitt R.N, The rice immune receptor XA21 recognizes a tyrosine-sulfated protein from a Gram-negative bacterium, *Science advances*, 1 (6), 2015, 1500245.
- Ronald P.C and Beutler B, Plant and animal sensors of conserved microbial signatures, *Science*, 330 (6007), 2010, 1061-1064.
- Ronald P.C, The Role of RaxST, a Prokaryotic Sulfotransferase, and RaxABC, a Putative Type I Secretion System, in Activation of the Rice XA21-Mediated Immune Response, *Scientifica*, 2014.
- Schurch S, Molecular population genetic analysis differentiates two virulence mechanisms of the fungal avirulence gene NIP1, *Molecular Plant-Microbe Interactions*, 17 (10), 2004, 1114-1125.
- Shen Y, The *Xanthomonas oryzae* pv. *oryzae* raxP and raxQ genes encode an ATP sulphurylase and adenosine-5'-phosphosulphate kinase that are required for AvrXa21 avirulence activity, *Molecular microbiology*, 44 (1), 2002, 37-48.
- Song W.Y, A receptor kinase-like protein encoded by the rice disease resistance gene, Xa21. *Science*, 270 (5243), 1995, 1804-1806.

Sun Y, Structural basis for flg22-induced activation of the Arabidopsis FLS2-BAK1 immune complex. *Science*, 342 (6158), 2013, 624-628.

Swings J, Reclassification of the Causal Agents of Bacterial Blight (*Xanthomonas campestris* pv. *oryzae*) and Bacterial Leaf Streak (*Xanthomonas campestris* pv. *oryzicola*) of Rice as Pathovars of *Xanthomonas oryzae* (ex Ishiyama 1922) sp. nov., nom. rev. *International Journal of Systematic Bacteriology*, 40 (3), 1990, 309-311.

Tao Y, Quantitative nature of Arabidopsis responses during compatible and incompatible interactions with the bacterial pathogen *Pseudomonas syringae*, *The Plant Cell*, 15 (2), 2003, 317-330.

Thomma B.P, Nurnberger T and Joosten M.H, of PAMPs and effectors, the blurred PTI-ETI dichotomy, *The plant cell*, 23 (1), 2011, 4-15.

Tsuda K and Katagiri F, Comparing signaling mechanisms engaged in pattern-triggered and effector-triggered immunity, *Current opinion in plant biology*, 13 (4), 2010, 459-465.

Tsuda K, Network properties of robust immunity in plants, *PLoS Genet*, 5 (12), 2009, 1000772.

Wang G.L, The cloned gene, Xa21, confers resistance to multiple *Xanthomonas oryzae* pv. *oryzae* isolates in transgenic plants. *Mol Plant Microbe Interact*, 9 (9), 1996, 850-855.

Wang Y.S, Rice XA21 binding protein 3 is a ubiquitin ligase required for full Xa21-mediated disease resistance, *The Plant Cell*, 18 (12), 2006, 3635-3646.

Xu W.H, The autophosphorylated Ser686, Thr688, and Ser689 residues in the intracellular juxtamembrane domain of XA21 are implicated in stability control of rice receptor-like kinase, *The Plant Journal*, 45 (5), 2006, 740-751.

Zipfel C, Plant pattern-recognition receptors, *Trends in immunology*, 35 (7), 2014, 345-351.